

final report

Project code:	A.MFS.0248			
Prepared by:	Darian Warne			
	DWC Food Tech Pty Ltd			
Date published:	January 2011			

PUBLISHED BY Meat & Livestock Australia Limited Locked Bag 991 NORTH SYDNEY NSW 2059

Low temperature cooking of meats

Meat & Livestock Australia acknowledges the matching funds provided by the Australian Government and contributions from the Australian Meat Processor Corporation to support the research and development detailed in this publication.

This publication is published by Meat & Livestock Australia Limited ABN 39 081 678 364 (MLA). Care is taken to ensure the accuracy of the information contained in this publication. However MLA cannot accept responsibility for the accuracy or completeness of the information or opinions contained in the publication. You should make your own enquiries before making decisions concerning your interests. Reproduction in whole or in part of this publication is prohibited without prior written consent of MLA.

Contents

1.	Background to the project	2
2.	Project objectives	2
3.	Methodology	3
4.	Results and Discussion	6
5.	Recommendations	8
6.	Appendix I	. 10

1. Background to the project

The introduction to the project brief received from MLA states that "Cooked meats that are ready to eat or are to be consumed with minimal reheating are susceptible to contamination with *Listeria monocytogenes*, which may grow in the product during (its) shelf-life and render the product unsafe for human co nsumption. Tables have been de veloped with time-temperature requirements for cooking to deliver to product a 6 D (log reduction) in L. *monocytogenes*. These tables are over 20 years old and do not exten d to the full range of temperatures that are used in manufacturing and food service operations."

The purpose of the project is to develop a table that shall be applicable for tempe ratures ranging from 55 to 85 °C. The table that shall be generated shall specify the exposure times at the SHP (slowest heating point) of cooked meats required to achieve, at least, a 6 log reduction in *L. monocytogenes*. The table will be based on data obtained from a literature review of the heat resistance of *L. monocytogenes* in a variety of meat and other products.

2. Project objectives

The objectives of the project are as follows:

- Review the scientific basis for time-temperature requirements in current food regulation. This shall include review of the codes issued by AQIS and USFDA and, where applicable, Good Manufacturing Practice recommendations from relevant advisory bodies and research organisations.
- 2. Review recent data and approaches to determining time-temperature requirements for safety in cooked meat products
- 3. Prepare time-temperature tables over the temperature range of 55-85°C.

3. Methodology

The papers and other publication s that were reviewed are cited in full in Appendix I and in abbreviated form beneath Table 1.

In order to determine the hold t ime required a t a spe cified temperature to bring a bout the desired 6 log (6*D*) reduction of the population of *L. monocytogenes* it is necessary to know each of the following,

- The *D* value, or the decimal reduction time, which is the time required at a specified reference temperature to bring about a 90% (i.e. a one log) reduction in a pure culture of the target microorganism (in this case *L. monocytogenes*).
- The *z* value of the target microorganism which is the number of degrees required to bring about a 10 fold change in its *D* value.
- The lethal rate L (at temperatures from 55 to 85 °C) relative to the lethal rate being unity at the reference temperature.

The equation for calculation of lethal rate is

	L	=	$Log^{-1} (T - T_r) / z$	(1)
where	Т	=	product temperature at the the core)	e slowest heating point (i.e. at
	T _r =		reference temperature	

z = number of degrees required to bring about a tenfold change in the decimal reduction time (*D*)

Shown in Table 1 are t he reference temperatures, *D* values, *z* values and the te st media (where cited) that were quoted in the nine sources that were reviewed. Using these values the lethal rates (L values) for each of the nine data sets were calculated over the 55 to 85 °C temperature range.

Once L values had be en determined using e quation 1, t he *D* values at t he no minated temperatures were then calculated using equation 2.

$$D_{\rm t} = D_{\rm ref} / L_{\rm t}$$
 (2)

where

- $D_{\rm t}$ = the *D* value at the nominated temperature t (between 55 and 85 °C)
- D_{ref} = the *D* value at the quoted reference temperature
- Lt = the lethal rate at the nominated temperatu re t (as calculated using equat ion 1) relat ive to the lethal rate being unity at the reference temperature

Once D_t values had been determined using equation 2, the hold time s required to bring about a six log reduction (i.e. a 6 *D* reduction) in *L. monocytogenes* at each of the core product temperatures between 55 and 85 °C were calculated. The products of these calculations have been presented in the body of Table 1.

When interpreting the data in Table 1 it is important to understand that the time-temp erature combinations shown within each of the eight sets of data are equivalent in lethality. This means, for i nstance, that each of the time-temperature combinations shown in the second column of T able 1 (i.e. Gaze *et al*¹) will bring about a 6 log reduction in *L. monocytogenes* counts. Similarly, with the other sev en sets of d ata, the time-temperature combinations for each will bring about 6 log reductions in *L. monocytogenes* counts.

It should also be understood that each of the time-tempe rature comb inations in Table 1 express the integrated lethality of a thermal treatment in terms of time at a reference temperature. This means that according to Jemmi and Stephan, and USFDA, 2.0 minutes at 70 °C is the total lethality of the process expressed in minutes at 70 °C even tho ugh the heating profile obtained by placing thermocouples at the slowest heating point of the product and recording temperature throughout the entire process (including cooling) may have shown that the core temperature did not reach, or may have exceed ed, 70 °C. Had the heating profile been based upon a reference temperature of 64 °C the total integrated process lethality would have been equivalent to exposure at 64 °C for 12.60 min.

Reference	Gaze et al ¹	Gaze et al ¹	ICMSF ²	AQIS ³ ,	Jemmi & Stephan ^⁵ ,	Doyle <i>et al</i>	Selby ⁸	Vanderlinde
				Cox & Bauler ⁴	USFDA ⁶			& Duffy ⁹
Reference temp (°C)	70	64	70	70	64	60	55	6
D value (min)	0.20	2.21	0.27	0.34	2.10	4.47	112.30	2.1
z value (C°)	5.98	5.98	7.50	7.50	7.50	7.90	5.50	8.0
Test medium	Beef steak	Beef steak	Meat slurry	-	-	Beef	Beef bologna	
Core temperature (°C)								
55	386.85	424.19	162.00	202.20	199.70	115.18	673.80	173.3
56	263.22	288.63	119.17	148.75	146.91	86.06	443.32	130.0
57	179.10	196.39	87.67	109.42	108.07	64.30	291.67	97.5
58	121.86	133.63	64.49	80.50	79.50	48.04	191.90	73.1
59	82.92	90.92	47.44	59.22	58.48	35.90	126.26	54.8
60	56.42	61.86	34.90	43.56	43.02	26.82	83.07	41.1
61	38.39	42.09	25.68	32.05	31.65	20.04	54.65	30.8
62	26.12	28.64	18.89	23.57	23.28	14.97	35.96	23.1
63	17.77	19.49	13.89	17.34	17.13	11.19	23.66	17.3
64	12.09	13.26	10.22	12.76	12.60	8.36	15.57	13.0
65	8.23	9.02	7.52	9.39	9.27	6.25	10.24	9.7
66	5.60	6.14	5.53	6.90	6.82	4.67	6.74	7.3
67	3.81	4.18	4.07	5.08	5.02	3.49	4.43	5.4
68	2.59	2.84	2.99	3.74	3.69	2.60	2.92	4.1
69	1.76	1.93	2.20	2.75	2.71	1.95	1.92	3.0
70	1.20	1.32	1.62	2.02	2.00	1.45	1.26	2.3
71	0.82	0.90	1.19	1.49	1.47	1.09	0.83	1.7
72	0.56	0.61	0.88	1.09	1.08	0.81	0.55	1.3
73	0.38	0.41	0.64	0.80	0.80	0.61	0.36	0.9
74	0.26	0.28	0.47	0.59	0.58	0.45	0.24	0.7
75	0.18	0.19	0.35	0.44	0.43	0.34	0.16	0.5
76	0.12	0.13	0.26	0.32	0.32	0.25	0.10	0.4
77	0.08	0.09	0.19	0.24	0.23	0.19	0.07	0.3
78	0.06	0.06	0.14	0.17	0.17	0.14	0.04	0.2
79	0.04	0.04	0.10	0.13	0.13	0.11	0.03	0.1
80	0.03	0.03	0.08	0.09	0.09	0.08	0.02	0.1
81	0.02	0.02	0.06	0.07	0.07	0.06	0.01	0.1
82	0.01	0.01	0.04	0.05	0.05	0.04	0.01	0.0
83	0.01	0.01	0.03	0.04	0.04	0.03	0.01	0.0
84	0.01	0.01	0.02	0.03	0.03	0.02	0.00	0.0
85	0.00	0.00	0.02	0.02	0.02	0.02	0.00	0.0

Table 1. Hold times (min) at product core temperatures of between 55 and 85 °C required to deliver 6 log reductions in Listeria monoctyogenes counts

References cited in Table 1.

- 1. Gaze et al. (1989).
- 2. ICMSF. (1996).
- 3. AQIS (1992).
- 4. Cox and Bauler (2008).
- 5. Jemmy and Stephan (2006).
- 6. US Food and Drug Administration (2001).
- 7. In Doyle et al. (2001).
- 8. Selby *et al.* (2006).
- 9. Vanderlinde and Duffy (1999).

4. Results and Discussion

Experimentally determined *D* values (and therefore, so-called, 6*D* processes) established under one set of condit ions, frequently are compared with t hose deter mined by other laborat ories under different conditions. However, several authors have commented that there can be limitations when making su ch direct comparisons and the reasons for this include the following:

- The thermal characteristics (*D* and *z* values) of *L. monocytogenes* are affected by the medium in which the microorganisms are heated (ICMSF, 1996; US Food and Drug Administration, 2001).
- The *D* value of *L. monocytogenes* is affected by 'strain v ariation, previous growth condit ions, exposure to heat shock, acid, and other stresses, and composition of the heating menstruum' (Doyle *et al.*, 2001).
- Gaze *et al* (1989) noted that 'the techniques used by differ ent groups of workers', including the container in which the microorganisms were heated, can affect the heat resistance of *L. monocytogenes.*
- Vanderlinde and Duff y (1999) observed that 'It is possible to extrapol ate the time /temperature combinations (requir ed by different countrie s) using these (diffe rent) z values, althoug h extrapolations can result in errors especially at temperatures well away from the da ta (the 60 to 76 °C range) presented.' This means, for example, that the 6 *D* hold time of 673.8 min at 55 °C (shown in Table 1 and based on the data in the paper by Selby *et al.* [2006].) ma y not b e accurately extrapolated to a 6*D* hold time of 0.01 min (0.6 s) at 83 °C.
- Different interpretations of data can lead to extrapolation of different conditions required to deliver 6 log reduct ions in *L. monocytogenes* counts. For example, the time-temperature combinations derived from the paper by Vanderlinde and Duffy (1999) and shown in Table 1 of t his report are not only conservative but also they highlight the inconsistencies that can occur when interpreting primary dat a. In Appen dix 1 of the ir paper Va nderlinde and Duffy in clude 6 *D* treatments for

L. monocytogenes of 12 min 45 s and 2 min 1 s at 64 and 70 °C, respectively. These values are in agreement with those shown in Table 1, according to AQIS (1992) and Cox and Bauler (2008) who use a z value of 7.5 °C. The 2 min 1 s at 70 °C, as quoted in the Appendix 1 of Vanderlinde and Duffy's (1999) paper, is effectively identical to the conservative 2 min at 70 °C value which is recommended by Gaze *et al* (1989). However, when using a z value of 8 °C (as shown in the body of Va nderlinde and Duffy's paper and in Table 1) the equivalent processes for a 6 log reduction of *L. monocytogenes* become 13.00 min (13 min 0 s) and 2.31 min (2 min 19 s) at 6 4 and 70 °C, respectively.

It is because of the individual and/or cumulative influence that the factors described above may have on the accuracy of determining minimum heating conditions for *L. monocytogenes* that caution should be exercised when extrapolating 6 D values to the second d ecimal place. For example, whilst the dat а presented by Gaze et al (1989) showed that the exposure times a t 70 °C re quired t o de liver 6 Dprocesses for *L. monocytogenes* ranged from 0.84 to 1.62 min when heating in homogenates containing steak, chicken or carrot, these authors nevertheless recommended 'that the slowest heating point in a product sho uld be held at 70 °C for 2 min t o ensure that it is eff ectively decontaminated of L. level of 10⁶ per co *monocytogenes* if cont amination is up to a ntainer.' In this insta nce the recommendation has been conservative and in corporates a 23% incre ase above the maximu m of the experimentally determined heating times in order to accommodate L. monocytogenes strains that might exhibit atypical D values under some test conditions.

5. Recommendations

In consideration of the inconsistencies that are apparent when comparing data from different studies and the different interpretations of those data, it is recommended that the heat treatments required to deliver 6 log reductions in *L. monocytogenes* should be equivalent to those shown in Table 2.

Temperature	Tir	ne
(°C)	(min)	(sec)
55	199	42
56	146	55
57	108	4
58	79	30
59	58	29
60	43	1
61	31	39
62	23	17
63	17	8
64	12	36
65	9	16
66	6	49
67	5	1
68	5 3 2 2 1	41
69	2	43
70	2	0
71	1	28
72	1	5
73	0	48
74	0	35
75	0	26
76	0	19
77	0	14
78	0	10
79	0	8
80	0	5
81	0	4
82	0	3
83	0	2
84	0	3 2 2 1
85	0	1

Table 2. Hold times¹ at product core temperatures of between 55 and 85 °C required to deliver 6 log reductions in*Listeria monoctyogenes* counts

1. Based on a D_{64} value of 2.10 min and a z value of 7.5 ${C}$

Points to note in interpretation of the data in Table 2 include the following:

- 1. The time/temperature combinations in Table 2 are all equ ivalent processes with respect to t he destruction of *L. monocytogenes*. For instance, assuming instantaneous heating and cooling at the slowest heating point of the product, a treatment t that maintains the core temperature at 60 °C for exactly 43 min and 1 second is equivalent with respect to the destruction of *L. monocytogenes* to a heat treatment at 70 °C for 2 min.
- 2. In practice, in commercial rather than laboratory applications, because of heating and cooling lags throughout the product mass, heating and cooling at the slowest heating point will not be instantaneous. For this reason the equivalence of the heat process is determined by integration of the time-temperature profile at the slowest heating point using a known reference temperature and a known z value.
- 3. The hold times shown in Table 2 a re those ne cessary at constant cor e (product) t emperatures in order to achieve 6 log reductions of *L. monocytogenes*. These hold times should not be confused with the actual process temperatures experienced in the heating vessel (e.g. in the hot water bath, or pasteurisation chamber, or hot air oven, or retort) which may e xhibit heating and cooling gradients during a thermal process.

As a hypothetical example consider the following. Heat penetration trials have shown that to deliver a 6 log reduction in *L. monocytogenes* counts in a 500 g pack of diced beef it is necessary to fully immerse the pack for 58 min in a water bath at 65 °C. In this example analysis of the he at penetration data shows that this process will deliver a pasteurising effect which is equivalent to the recommended 2 min at 70 °C. Calculations also show that equivalent processes could have been achieved by immersion at 55 °C for 250 min, or 74 °C for 38 min. This means that each of these three hypothetical processes is equivalent to 2 min at 70 °C at the s lowest heating point of the pack.

- 4. In order to satisfy the recommendations shown in Table 2 and establish a safe thermal process it is necessary t o conduct replicate h eat penetra tion tria ls, under worst-case conditions, using calibrated t emperature logging equipment. As a guide it is recomme nded that at least six h eat penetration data sets are gathered.
- 5. To comply with Good Manufacturing Practice it is recommended that temperature distribution trials throughout the heating vessel are conducted in order to determine whe ther and where cold sp ots may be located. Unless identified cold spots can lead to delivery of insufficient thermal processes.

6. Appendix I

References.

- 1. Gaze, J. E., Brown, G. D., Gaskell, D. E. and Banks, J. G. (1989). *Heat resistance of <u>Listeria</u>* <u>monocytogenes</u> in homogenates of chicken, beef steak and carrot. Food Microbiology 6: 251 - 259.
- 2. ICMSF. (1996). *Microorganisms in Foods 5. Characteristics of Microbial Pathogens*. Blackie Academic and Professional, London, pp. 141-182.
- AQIS, Australian Quarantine and Inspection Service, Department of Primary Industries & Energy. (1992). Code of hygienic practice for heat-treated refrigerated foods packaged for extended shelflife. Australian Government Publishing Service, Canberra.
- 4. Cox B and Bauler, M. (2008). *Cook chill for foodservice and manufacturing: Guidelines for safe production, storage and distribution*. AIFST, Australian Institute of Food Science and Technology Inc. Alexandria, NSW, pp 25 and 157.
- 5. Jemmi, T. and Stephan, R. (2006). *Listeria monocytogenes: food-borne pathogen and hygiene indicator*. Rev. Sci. tech. Off. Int. Epiz., 25 (2): 571 580.
- 6. US Food and Drug Administration. Fish and Fisheries Products Hazards and Controls Guidance Third Edition (2001) in Appendix 4 - Bacterial Pathogen Growth and Inactivation. Available at <u>http://www.fda.gov/Food/GuidanceComplianceRegulatoryInformation/GuidanceDocuments/Seafoo</u> <u>d/FishandFisheriesProductsHazardsandControlsGuide/ucm120106.htm</u>. Last updated 11/10/2009.
- Doyle, M. E., Mazzotta, A. S., Wang, T., Wiseman, D. W. and Scott, V. N. (2001). *Heat resistance of <u>Listeria monocytogenes</u>*. Journal of Food Protection, 64 (3): 410 429.
- Selby, T. L., Berzins, A., Gerrard, D. E., Corvalen, C. M., Grant, A. L. and Linton, R.H. (2006). Microbial heat resistance of <u>Listeria monocytogenes</u> and the impact of ready-to-eat meat quality after post-package pasteurization. Meat Science 74: 425 – 434.
- 9. Vanderlinde, P. and Duffy, L. (1999). Thermal destruction of bacteria. National and International Standards. Food Science Australia, Cannon Hill.
- 10. Warne, D. (2005). *Technological developments and safety of heat processed foods*. PhD Thesis, University of Tasmania.